

5th MMTN Conference 5-6 November 2016

Bangkok, Thailand

10:20-10:45, 6 Nov, 2016

# **Antifungal Resistance in Asia: Mechanisms, Epidemiology, and Consequences**

---

**Yee-Chun Chen, M.D., PhD.**

Department of Medicine, National Taiwan University  
Hospital and College of Medicine; National Institute of  
Infectious Diseases and Vaccinology, National Health  
Research Institutes, Taiwan





# Disclosure

---

- Research grants: Pfizer, Gilead
- Advisory boards: Pfizer, MSD, Gilead
- Speaker: Pfizer, MSD, Gilead

# Contents

---

- What is meant by resistance?
- Is antifungal resistance in *Candida* and other yeasts a problem in Asia?
- Is antifungal resistance in *Aspergillus* and other molds a problem in Asia?
- Limitation of current published data
- Conclusion



# What is meant by resistance?

**Clinical failure**

**failure of drug therapy to resolve signs and symptoms of infection**

**Microbiological failure**

**failure of therapy to eradicate the organism**

**In vitro definition**

**based on minimum inhibitory concentrations and clinical breakpoints**

# CLSI clinical breakpoints of antifungal resistance against *Candida* species

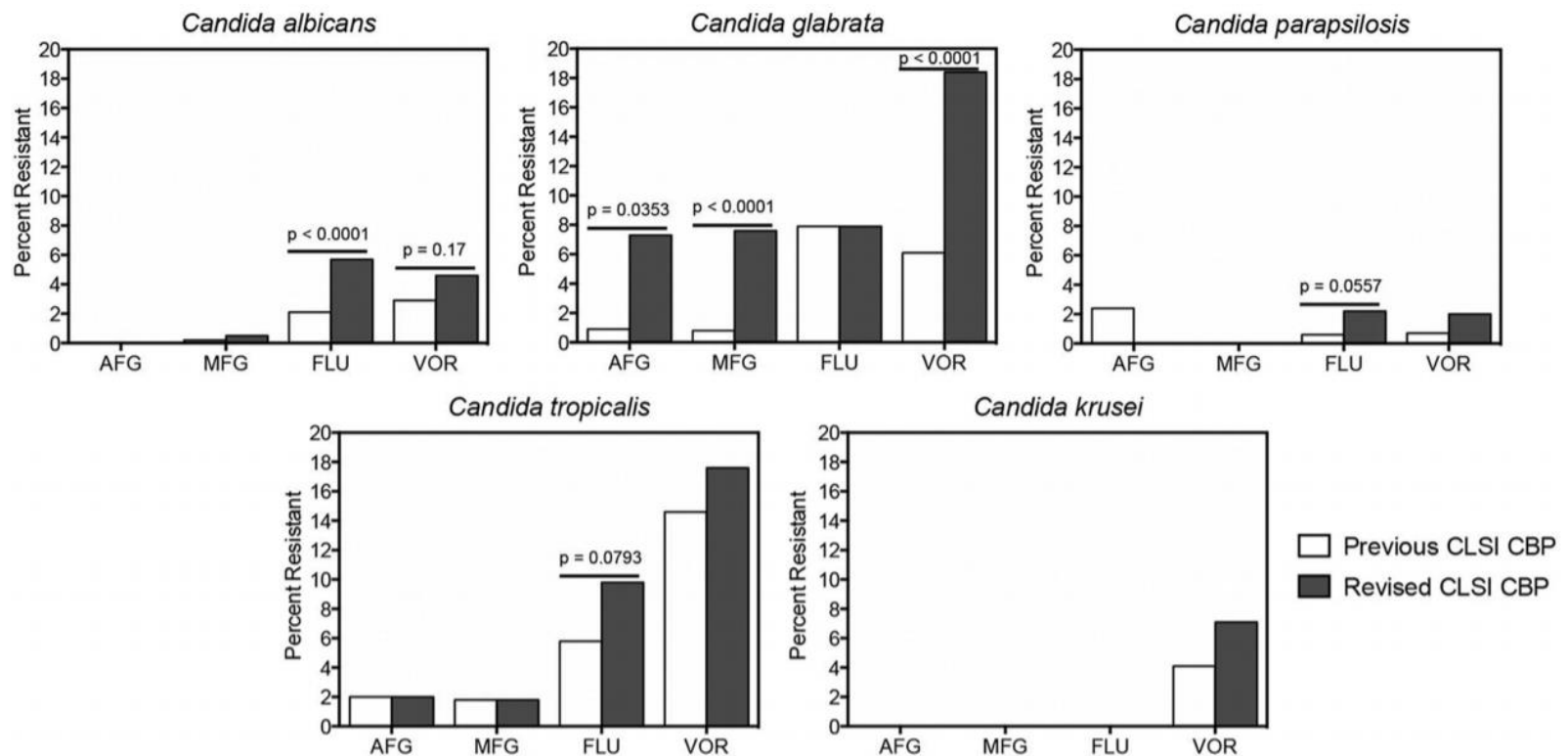
		<i>C. albicans</i>	<i>C. glabrata</i>	<i>C. tropicalis</i>	<i>C. krusei</i>	<i>C. parapsilosis</i>
Anidulafungin	Old	≥4	≥4	≥4	≥4	≥4
	New	≥1	≥0.5	≥1	≥1	≥8
Micafungin	Old	≥4	≥4	≥4	≥4	≥4
	New	≥1	≥0.25	≥1	≥1	≥8
Fluconazole	Old	≥64	≥64	≥64	≥64	≥64
	New	≥8	≥64	≥8	≥8	≥8
Voriconazole	Old	≥4	≥4	≥4	≥4	≥4
	New	≥1	≥1 *	≥1	≥2	≥1

\*epidemiologic cutoff value

Clinical and Laboratory Standards Institute (CLSI) **2008**, M27-A3; **2012**. M27-S4.



# Impact of New Antifungal Breakpoints on Antifungal Resistance in *Candida*



# EUCAST Antifungal Clinical Breakpoint – Yeasts

v. 8.0 valid from 2015-11-16

Antifungal agent	MIC breakpoint (mg/L)													
	<i>C. albicans</i>		<i>C. glabrata</i>		<i>C. krusei</i>		<i>C. parapsilosis</i>		<i>C. tropicalis</i>		<i>C. guilliermondii</i>		Non-species related breakpoints <sup>1</sup>	
	S ≤	R >	S ≤	R >	S ≤	R >	S ≤	R >	S ≤	R >	S ≤	R >	S ≤	R >
Amphotericin B	1	1	1	1	1	1	1	1	1	1	IE	IE	IE	IE
Anidulafungin	0.03	0.03	0.06	0.06	0.06	0.06	0.002	4	0.06	0.06	IE <sup>2</sup>	IE <sup>2</sup>	IE	IE
Caspofungin	Note <sup>3</sup>	Note <sup>3</sup>	Note <sup>3</sup>	Note <sup>3</sup>	Note <sup>3</sup>	Note <sup>3</sup>	Note <sup>3</sup>	Note <sup>3</sup>	Note <sup>3</sup>	Note <sup>3</sup>	IE <sup>2</sup>	IE <sup>2</sup>	IE	IE
Fluconazole	2	4	0.002	32	-	-	2	4	2	4	IE <sup>2</sup>	IE <sup>2</sup>	2	4
Isavuconazole	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE
Itraconazole	0.06	0.06	IE <sup>2</sup>	IE <sup>2</sup>	IE <sup>2</sup>	IE <sup>2</sup>	0.12	0.12	0.12	0.12	IE <sup>2</sup>	IE <sup>2</sup>	IE	IE
Micafungin	0.016	0.016	0.03	0.03	IE <sup>4</sup>	IE <sup>4</sup>	0.002	2	IE <sup>4</sup>	IE <sup>4</sup>	IE <sup>4</sup>	IE <sup>4</sup>	IE	IE
Posaconazole	0.06	0.06	IE <sup>2</sup>	IE <sup>2</sup>	IE <sup>2</sup>	IE <sup>2</sup>	0.06	0.06	0.06	0.06	IE <sup>2</sup>	IE <sup>2</sup>	IE	IE
Voriconazole	0.12 <sup>5</sup>	0.12 <sup>5</sup>	IE	IE	IE	IE	0.12 <sup>5</sup>	0.12 <sup>5</sup>	0.12 <sup>5</sup>	0.12 <sup>5</sup>	IE <sup>2</sup>	IE <sup>2</sup>	IE	IE

# EUCAST Antifungal Clinical Breakpoint - Molds

v. 8.0 valid from 2015-11-16

Antifungal agent	MIC breakpoint (mg/L)											
	<i>A. flavus</i>		<i>A. fumigatus</i>		<i>A. nidulans</i>		<i>A. niger</i>		<i>A. terreus</i>		Non-species related breakpoints <sup>1</sup>	
	S ≤	R >	S ≤	R >	S ≤	R >	S ≤	R >	S ≤	R >	S ≤	R >
Amphotericin B	IE <sup>2</sup>	IE <sup>2</sup>	1	2	Note <sup>3</sup>	Note <sup>3</sup>	1	2	-	-	IE	IE
Anidulafungin	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE
Caspofungin	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE
Fluconazole	-	-	-	-	-	-	-	-	-	-	-	-
Isavuconazole	IE <sup>2</sup>	IE <sup>2</sup>	1	1	0.25	0.25	IE <sup>2</sup>	IE <sup>2</sup>	1	1	IE	IE
Itraconazole <sup>4</sup>	1	2	1	2	1	2	IE <sup>2,5</sup>	IE <sup>2,5</sup>	1	2	IE <sup>5</sup>	IE <sup>5</sup>
Micafungin	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE	IE
Posaconazole <sup>4</sup>	IE <sup>2</sup>	IE <sup>2</sup>	0.12 <sup>6</sup>	0.25 <sup>6</sup>	IE <sup>2</sup>	IE <sup>2</sup>	IE <sup>2</sup>	IE <sup>2</sup>	0.12 <sup>6</sup>	0.25 <sup>6</sup>	IE	IE
Voriconazole <sup>4</sup>	IE <sup>2</sup>	IE <sup>2</sup>	1	2	IE	IE	IE <sup>2</sup>	IE <sup>2</sup>	IE <sup>2</sup>	IE <sup>2</sup>	IE	IE



**Is antifungal resistance in  
*Candida* and other yeasts a  
problem in Asia?**

---

# Get ready for voting...

Go to webpage: [pollev.com/mmtn](http://pollev.com/mmtn)

Voting question on the next slide

Wifi network: MMTN2016  
Password: 2016mmtn



Q. Which is your most commonly used empirical antifungal agent for invasive candidiasis?

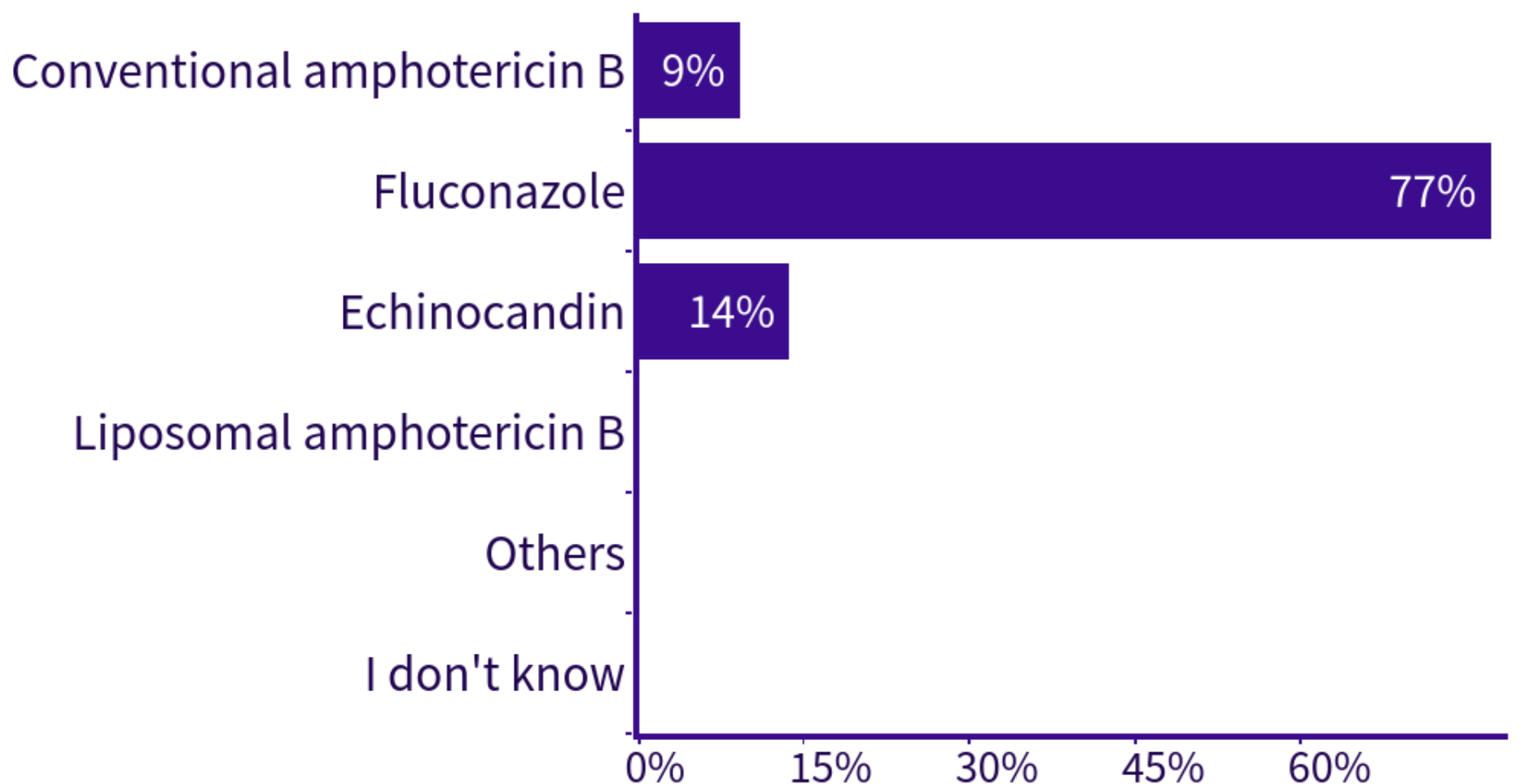
---

- A. Conventional amphotericin B
- B. Fluconazole
- C. Echinocandin
- D. Liposomal amphotericin B
- E. Others
- F. I don't know

# Which is your most commonly used empirical antifungal agent for invasive candidiasis?



When poll is active, respond at **PollEv.com/mmtn**



Total Results: 22



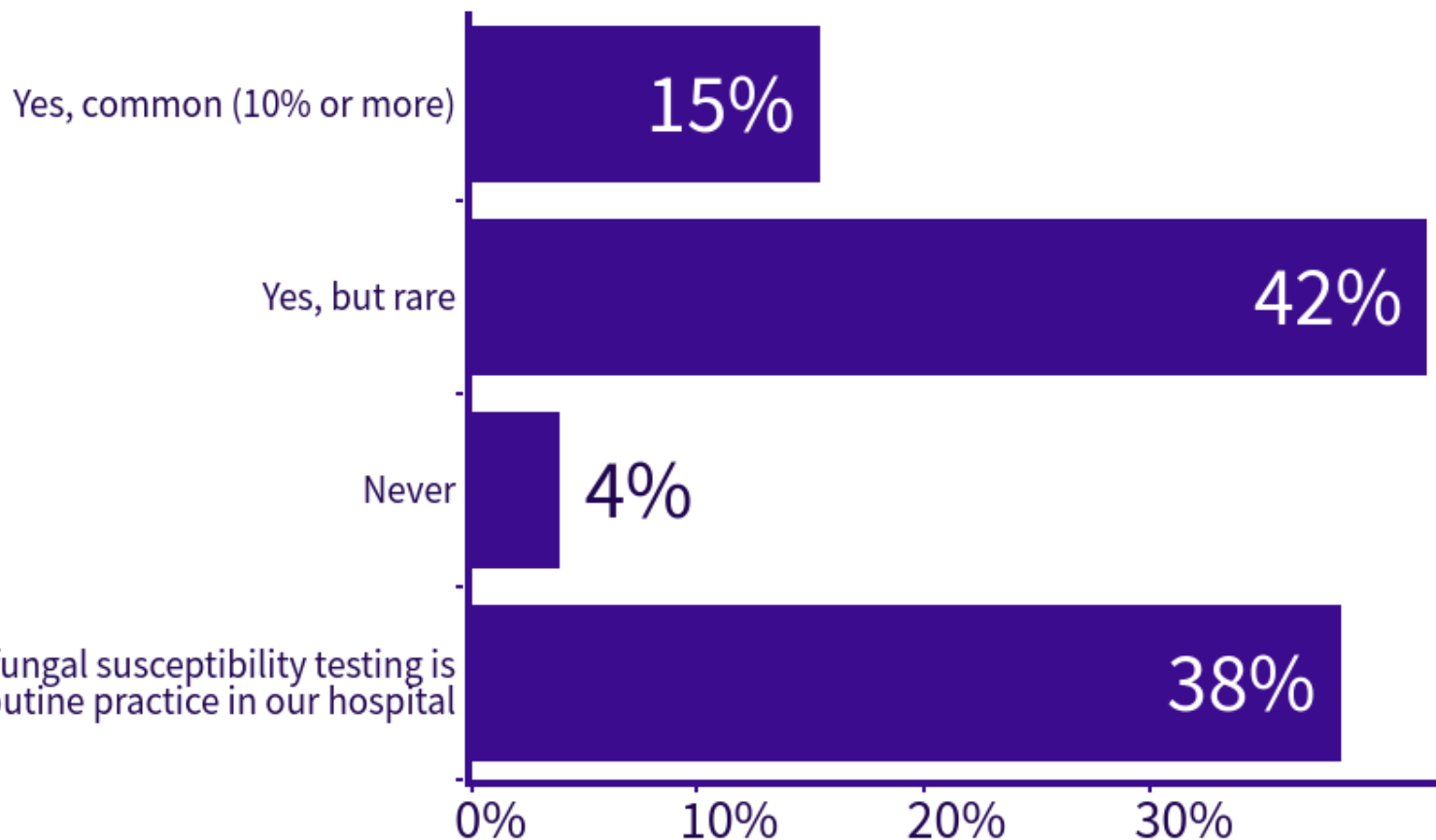
Q. Does antifungal resistance compromise the use of current empirical antifungal agent for invasive candidiasis in your hospital?

---

- A. Yes, common (10% or more)
- B. Yes, but rare
- C. Never
- D. I do not know as antifungal susceptibility testing is not available for routine practice in our hospital

# Does antifungal resistance compromise the use of current empirical antifungal agent for invasive candidiasis in your hospital?

**i** Poll is full and no longer accepting responses



Total Results: 26



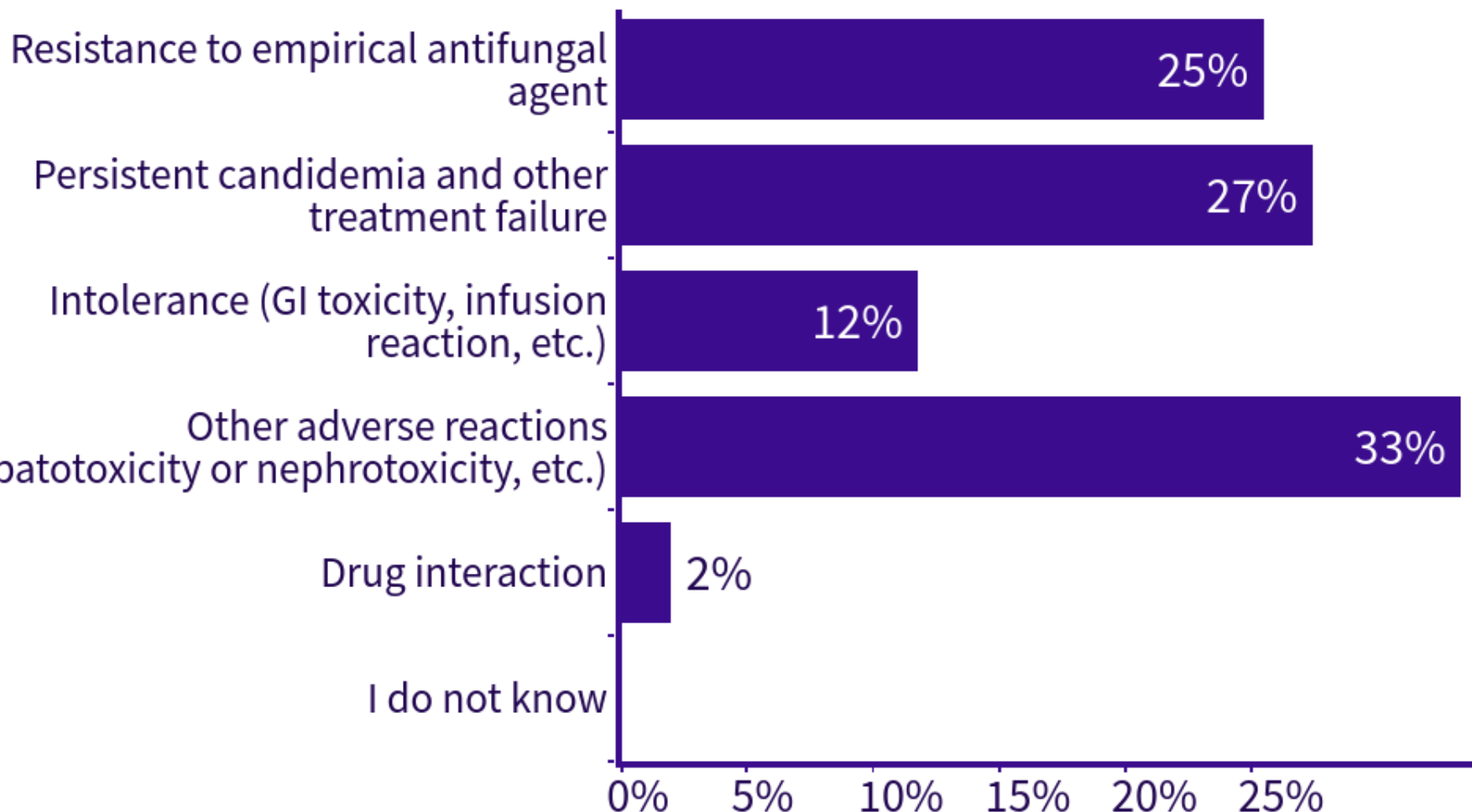
Q. The most common two reasons of modification of therapeutic agents for invasive candidiasis? (Select two choices)

---

- A. Resistance to empirical antifungal agent
- B. Persistent candidemia and other treatment failure
- C. Intolerance (GI toxicity, infusion reaction, etc.)
- D. Other adverse reactions (hepatotoxicity or nephrotoxicity, etc.)
- E. Drug interaction
- F. I do not know

# The most common two reasons of modification of therapeutic agents for invasive candidiasis? (Select two choices)

 Poll is full and no longer accepting responses



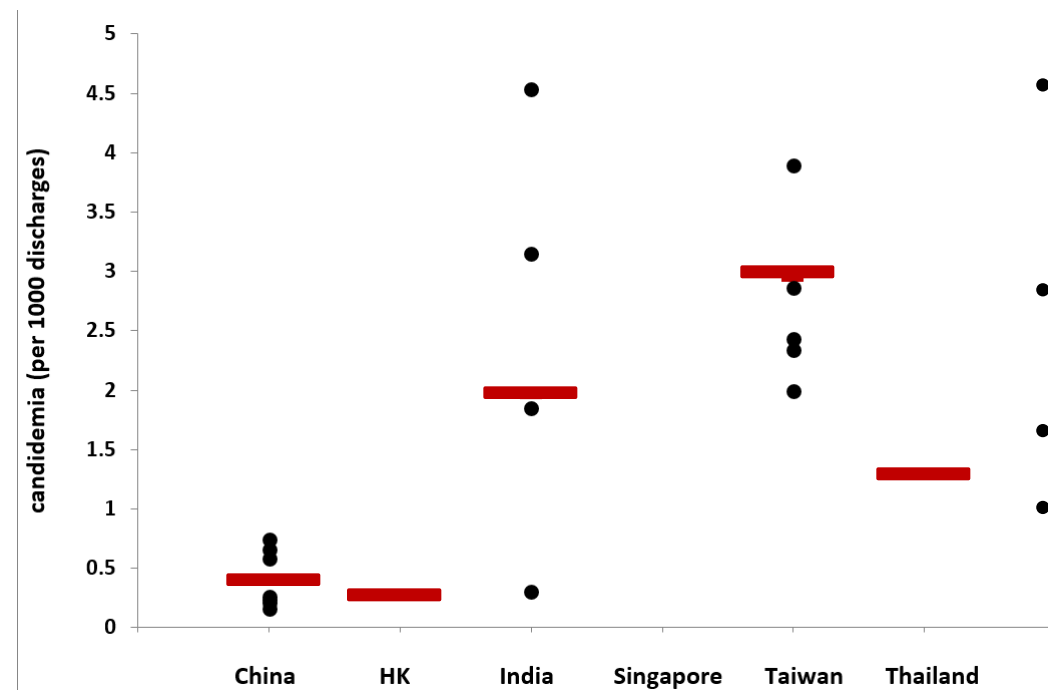
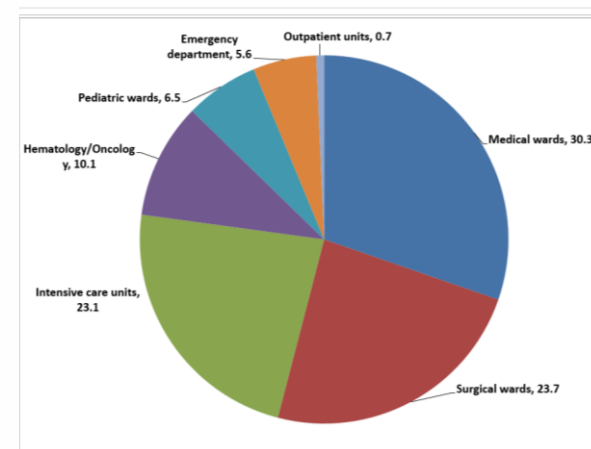
Total Results: 51



# Incidence and species distribution of candidaemia in Asia: a laboratory-based surveillance study

B. H. Tan<sup>1</sup>, A. Chakrabarti<sup>2</sup>, R. Y. Li<sup>3</sup>, A. K. Patel<sup>4</sup>, S. P. Watcharananan<sup>5</sup>, Z. Liu<sup>6</sup>, A. Chindamporn<sup>7</sup>, A. L. Tan<sup>8</sup>, P.-L. Sun<sup>9</sup>, U.-I. Wu<sup>10</sup> and Y.-C. Chen<sup>11,12</sup>, on behalf of the Asia Fungal Working Group (AFWG)

1) Department of Infectious Diseases, Singapore General Hospital, Singapore, 2) Department of Medical Microbiology, Postgraduate Institute of Medical Education & Research (PGIMER), Chandigarh, India, 3) Department of Dermatology, Peking University First Hospital, Research Centre for Medical Mycology, Peking University, Beijing, China, 4) Department of Infectious Diseases, Sterling Hospital, Ahmedabad, India, 5) Division of Infectious Disease, Department of Medicine, Faculty of Medicine, Ramathibodi Hospital, Bangkok, Thailand, 6) Department of Infectious Diseases, Peking Union Medical College Hospital, Beijing, China, 7) Department of Microbiology, Faculty of Medicine, King Chulalongkorn Memorial Hospital Chulalongkorn University, Bangkok, Thailand, 8) Department of Pathology, Singapore General Hospital, Singapore, 9) Department of Dermatology, Mackay Memorial Hospital, 10) Department of Medical Research, National Taiwan University Hospital, 11) Department of Medicine, National Taiwan University Hospital and College of Medicine, Taipei and 12) National Institute of Infectious Diseases and Vaccinology, National Health Research Institutes, Miaoli County, Taiwan

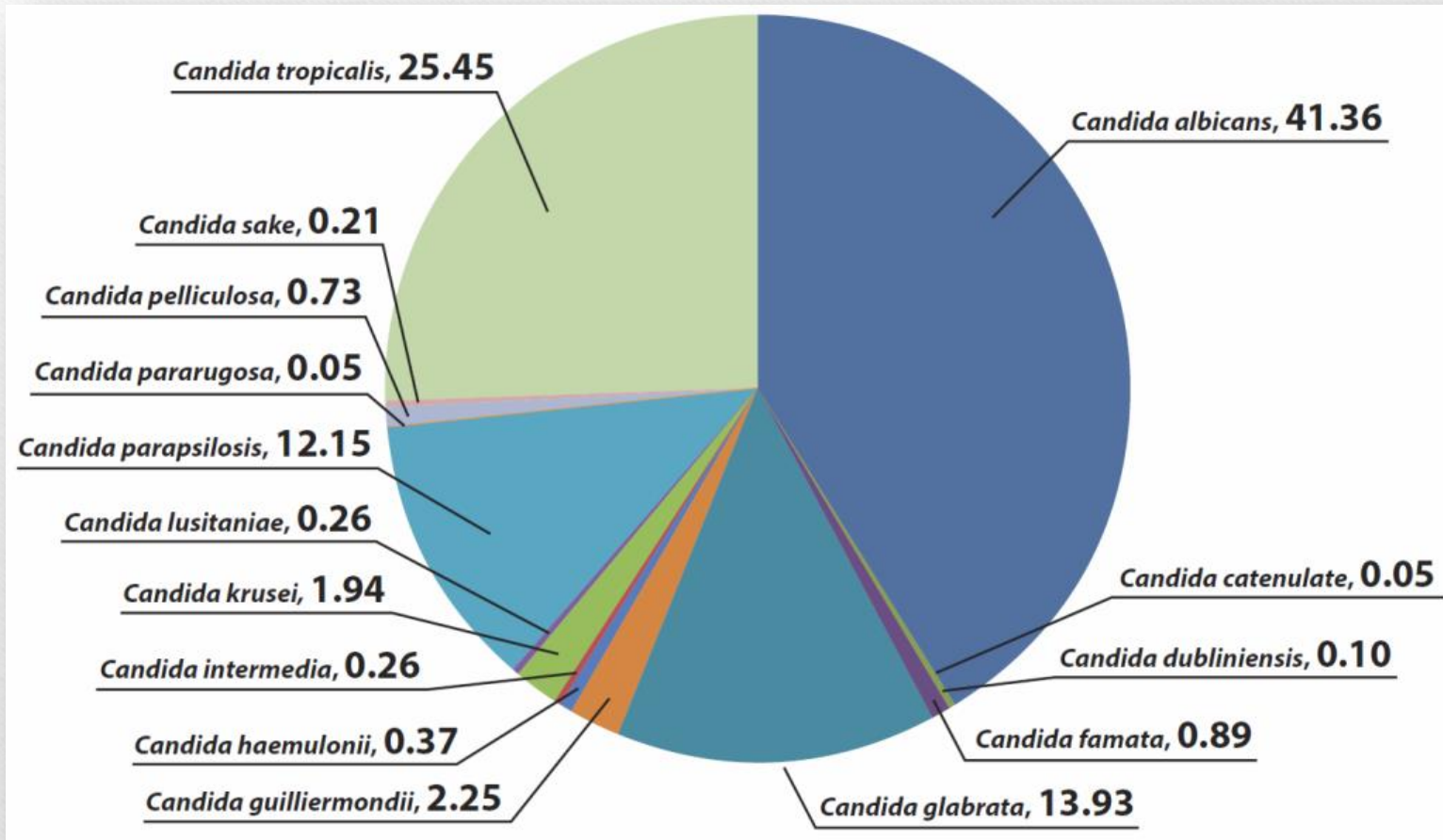


No. of hospitals	9	1	3	1	6	3
No. of episodes	310	30	330	73	1104	130

- The overall incidence was **1.22** episodes per 1,000 discharges or 0.15 episodes/1000 patient-days
- Varied among the hospitals and countries.
- ICU: **11.7** per 1000 discharges
- There was a moderate correlation between incidence of candidemia and the ICU/total bed ratio ( $R^2=0.47$ )

# ***Candida tropicalis* was the leading non-*albicans* species**

**Species distribution in Asia ,1910 non-duplicate blood *Candida* isolates**





## Both geographic and healthcare contribute to the variation of species distribution of candidemia

---

- *C. tropicalis* was more likely to be isolated at neutropenic patients than others (39% vs 17%).<sup>1</sup>
- *C. tropicalis* was more likely to be isolated at hemato-oncolgy wards than others (34.0% vs 24.5%).<sup>2</sup>
- The proportions of *C. tropicalis* among blood isolates were higher in tropical areas (India, Thailand and Singapore) than other geographical regions (46.2% versus 18.9%).<sup>2</sup>

1. Hung et al. J Formos Med Assoc 1996;95:19

2. Tan et al. Clin Microbiol Infect 2015; 21: 946

# *Candida* bloodstream isolates, Asia-Pacific region

Species	Antifungal name	%I	%S	MIC50 (mg/l)	MIC90 (mg/l)
<i>Candida albicans</i>	Anidulafungin	0	100	0.016	0.064
	Caspofungin	0	100	0.064	0.064
	Fluconazole*	0	99.7	0.25	0.5
	Micafungin	0	100	0.008	0.016
	Voriconazole*	0	100	0.008	0.008
<i>Candida tropicalis</i>	Anidulafungin	0.4	99.2	0.032	0.125
	Caspofungin	0.4	99.6	0.064	0.125
	Fluconazole*	6.1	75.8	2	32
	Micafungin	0	100	0.032	0.032
	Voriconazole*	16.7	69.3	0.125	1
<i>Candida parapsilosis</i>	Anidulafungin	0	100	0.5	2
	Caspofungin	0	100	0.25	0.5
	Fluconazole*	3.0	94.8	0.5	2
	Micafungin	0.8	99.2	1	2
	Voriconazole*	0.8	99.2	0.008	0.032
<i>Candida glabrata</i>	Anidulafungin	0	99.1	0.032	0.064
	Caspofungin	5.2	93.1	0.125	0.125
	Fluconazole*	94.8	n/a	8	32
	Micafungin	1.7	98.3	0.016	0.016

861 isolates from 13 centers, 2014

Sensititre YeastOne YST-010, Thermofisher, United Kingdom

Tan TY, et al. *Med Mycology* 2016;54, 471



# *Candida* bloodstream isolates, Asia-Pacific region

Species	Antifungal name	Brunei		Korea		Philippines		Taiwan		Thailand		Singapore		Vietnam	
		%I	%S	%I	%S	%I	%S	%I	%S	%I	%S	%I	%S	%I	%S
<i>Candida albicans</i>	Anidulafungin	0	100	0	100	0	100	0	100	0	100	0	100	0	100
	Caspofungin	0	100	0	100	0	100	0	100	0	100	0	100	0	100
	Fluconazole*	0	100	0	97.4	0	100	0	100	0	100	0	100	0	100
	Micafungin	0	100	0	100	0	100	0	100	0	100	0	100	0	100
	Voriconazole*	0	100	0	100	0	100	0	100	0	100	0	100	0	100
<i>Candida tropicalis</i>	Anidulafungin	0	100	0	100	0	100	0	96.6	0	100	2.3	97.7	0	100
	Caspofungin	0	100	0	100	0	100	0	100	0	100	2.3	97.7	0	100
	Fluconazole*	14.3	85.7	0	100	0	100	6.9	82.8	9.5	70.3	6.8	72.7	3.3	61.7
	Micafungin	0	100	0	100	0	100	0	100	0	100	0	100	0	100
	Voriconazole*	7.1	92.9	0	100	3.8	96.2	13.8	82.8	16.2	64.9	25	63.6	25	46.7
<i>Candida parapsilosis</i>	Anidulafungin	0	100	0	100	0	100	0	100	0	100	0	100	0	100
	Caspofungin	0	100	0	100	0	100	0	100	0	100	0	100	0	100
	Fluconazole*	11.1	83.3	3.8	96.2	4.2	87.5	9.0	91.0	0	100	0	95	0	100
	Micafungin	5.6	94.4	0	100	0	100	0	100	0	100	0	100	0	100
	Voriconazole*	0	100	0	100	4.2	95.8	0	100	0	100	0	100	0	100
<i>Candida glabrata</i>	Anidulafungin	0	100	0	100	0	100	0	100	0	96.8	0	100	0	100
	Caspofungin	0	100	22.2	77.8	0	100	0	100	3.2	93.5	8.2	89.8	0	100
	Fluconazole*	80	n/a	100	n/a	100	n/a	100	n/a	93.5	n/a	93.9	n/a	100	n/a
	Micafungin	0	100	0	100	0	100	0	100	3.2	96.8	2	98	0	100

# Invasive candidiasis in intensive care units in China

## the China-SCAN study

Species	Isolates, n (%)	Agent	MIC (mg/L)			Susceptible, n (%)	SDD, n (%)	Resistant, n (%)	ECV (mg/L)	
			range	50%	90%				WT	non-WT
<i>Candida albicans</i>	156 (40.1)	fluconazole	0.06–64	1	4	134 (85.9)	7 (4.5)	15 (9.6)	156 (100.0)	0 (0.0)
		voriconazole	0.03–0.5	0.03	0.03	145 (93.0)	11 (7.1)	0 (0.0)		
		itraconazole	0.06–4	0.06	1	6 (3.9)	125 (80.1)	25 (16.0)		
		caspofungin	0.03–0.25	0.06	0.125	156 (100.0)	0 (0.0)	0 (0.0)		
		amphotericin B	0.25–1	0.5	0.5					
<i>Candida parapsilosis</i>	83 (21.3)	fluconazole	0.125–64	4	8	40 (48.2)	27 (32.5)	16 (19.3)	50 (60.2)	33 (39.8)
		voriconazole	0.03–2	0.03	0.125	77 (92.8)	3 (3.6)	3 (3.6)		
		itraconazole	0.06–2	0.5	1					
		caspofungin	0.03–0.5	0.25	0.25	83 (100.0)	0 (0.0)	0 (0.0)		
		amphotericin B	0.25–1	0.5	0.5					
<i>Candida tropicalis</i>	67 (17.2)	fluconazole	0.25–64	2	4	42 (62.7)	21 (31.3)	4 (6.0)	46 (68.7)	21 (31.3)
		voriconazole	0.03–0.5	0.03	0.25	60 (89.6)	7 (10.4)	0 (0.0)		
		itraconazole	0.03–4	0.5	2					
		caspofungin	0.03–0.25	0.06	0.125	67 (100.0)	0 (0.0)	0 (0.0)		
		amphotericin B	0.25–1	0.5	1	67 (100.0) <sup>a</sup>		0 (0.0) <sup>a</sup>		
<i>Candida glabrata</i>	50 (12.9)	fluconazole	0.5–64	4	32	0 (0.0)	48 (96.0)	2 (4.0)	47 (94.0)	3 (6.0)
		voriconazole	0.03–2	0.06	0.125					
		itraconazole	0.5–4	1	2	0 (0.0)				
		caspofungin	0.03–0.25	0.125	0.25	43 (86.0)	7 (14.0)	0 (0.0)		
		amphotericin B	0.03–1	0.5	0.5	50 (100.0) <sup>a</sup>		0 (0.0) <sup>a</sup>		
<i>Candida haemulonii</i>	15 (3.9)	fluconazole	1–64	16	16	—		—		
		voriconazole	0.03–4	0.25	0.5	—		—		
		itraconazole	0.5–2	1	2	—		—		
		caspofungin	0.06–0.125	0.06	0.06	—		—		
		amphotericin B	0.5–1	0.5	1	—		—		



# ICU-acquired candidemia in India



27 Indian ICUs

Arunaloke Chakrabarti et al. Intensive Care Med (2015) 41:285

Antifungal	AFST	All species (n = 918)	<i>C. tropicalis</i> (n = 382)
Amphotericin B	MIC <sub>50</sub> (µg/ml)	–	0.50
	MIC <sub>90</sub> (µg/ml)	–	1.00
	Resistant (%)	2.1 %	4 (1.0)
	MIC percentile (25–75)	–	0.25–1
Fluconazole	MIC <sub>50</sub> (µg/ml)	–	0.50
	MIC <sub>90</sub> (µg/ml)	–	2.00
	Resistant (%)	6.2 %	10 (2.6)
	SDD (%)	11.0 %	9 (2.4)
Itraconazole	MIC percentile (25–75)	–	0.25–1
	MIC <sub>50</sub> (µg/ml)	–	0.06
	MIC <sub>90</sub> (µg/ml)	–	0.12
	Resistant (%)	1.2 %	1 (0.3)
Posaconazole	SDD (%)	9.3 %	27 (7.1)
	MIC percentile (25–75)	–	0.03–0.12
	MIC <sub>50</sub> (µg/ml)	–	0.03
	MIC <sub>90</sub> (µg/ml)	–	0.25
Voriconazole	MIC percentile (25–75)	–	0.03–0.12
	MIC <sub>50</sub> (µg/ml)	–	0.12
	MIC <sub>90</sub> (µg/ml)	–	0.50
	Resistant (%)	5.6 %	31 (8.1)
Anidulafungin	SDD (%)	22.9 %	128 (33.5)
	MIC percentile (25–75)	–	0.06–0.25
	MIC <sub>50</sub> (µg/ml)	–	0.03
	MIC <sub>90</sub> (µg/ml)	–	0.25
Caspofungin	Resistant (%)	1.7 %	8 (2.1)
	Intermediate (%)	1.6 %	8 (2.1)
	MIC percentile (25–75)	–	0.03–0.06
	MIC <sub>50</sub> (µg/ml)	–	0.25
Micafungin	MIC <sub>90</sub> (µg/ml)	–	0.50
	Resistant (%)	5.6 %	16 (4.2)
	Intermediate (%)	10.1 %	50 (13.1)
	MIC percentile (25–75)	–	0.12–0.25
	MIC <sub>50</sub> (µg/ml)	–	0.03
	MIC <sub>90</sub> (µg/ml)	–	0.12
	Resistant (%)	1.7 %	5 (1.3)
	Intermediate (%)	2.2 %	11 (2.9)
	MIC percentile (25–75)	–	0.03



# *C. tropicalis*

---

- The time-to-positivity (TTP) of blood cultures of *C. tropicalis* was significantly shorter than that of other species<sup>1</sup>
- Septic shock and skin emboli are common findings of candidemia<sup>2</sup>
- Both short TTP<sup>3</sup> and septic shock are associated with poor prognosis.
- A survey in Taiwan found genetically related *C. tropicalis* exhibiting reduced susceptibility to fluconazole from the human hosts and environmental samples.<sup>4</sup>

1. Lai et al. J Med Microbiol 2012;61:701

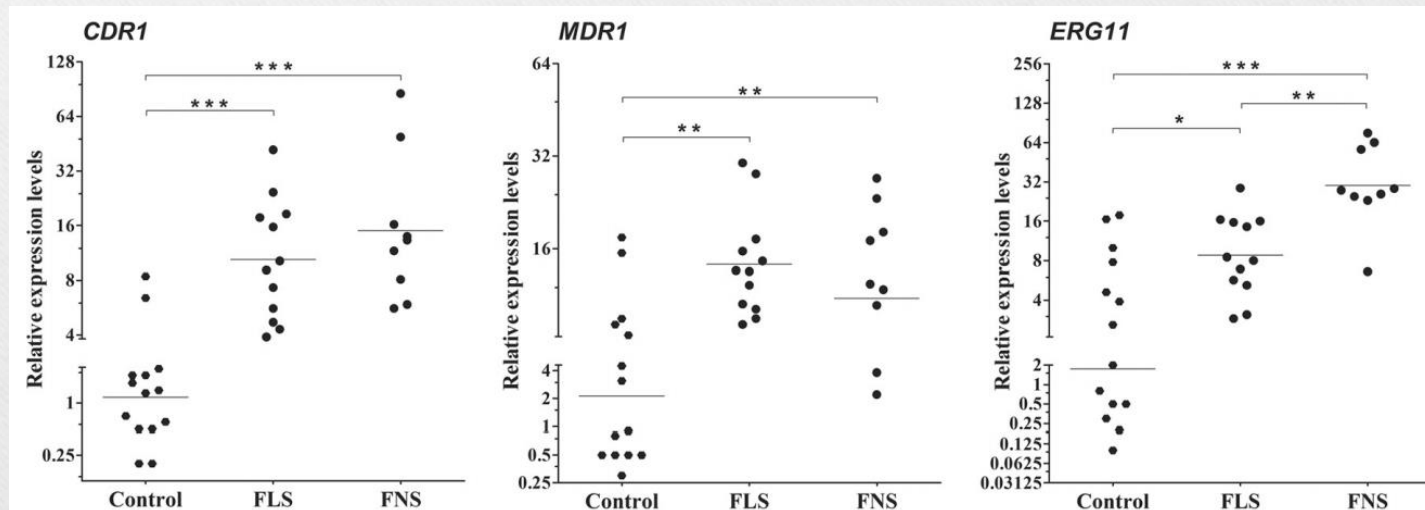
2. Leung et al. J Hosp Infect 2002;50:316

3. Kim et al. J Antimicrob Chemother 2013

4. Yang et al. PLoS One 2012;7:e34609.

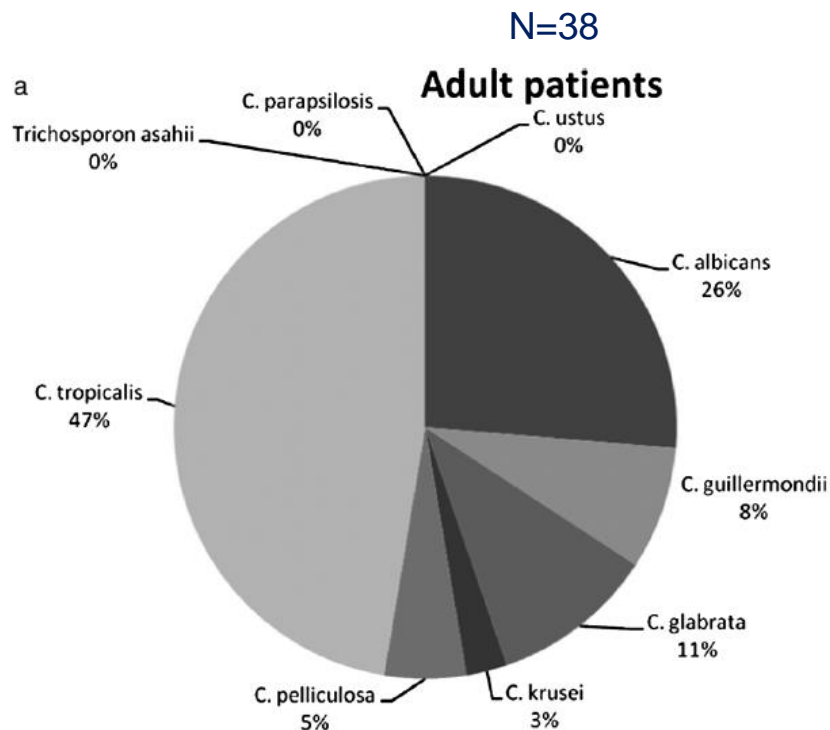
# Fluconazole-nonsusceptible/less-susceptible *C. tropicalis*

- FNS (MIC  $\geq 4$  g/ml), isolates were identified more frequently from patients with previous azole exposure (6/6 versus 3/10;  $P = 0.011$ ) and immunosuppression (6/6 versus 3/10;  $P = 0.011$ ).
- FNS and FLS (1-2 g/ml), bloodstream isolates were associated with azole therapeutic failure (3/4 versus 4/7) or uncleared fungemia (4/6 versus 4/10)



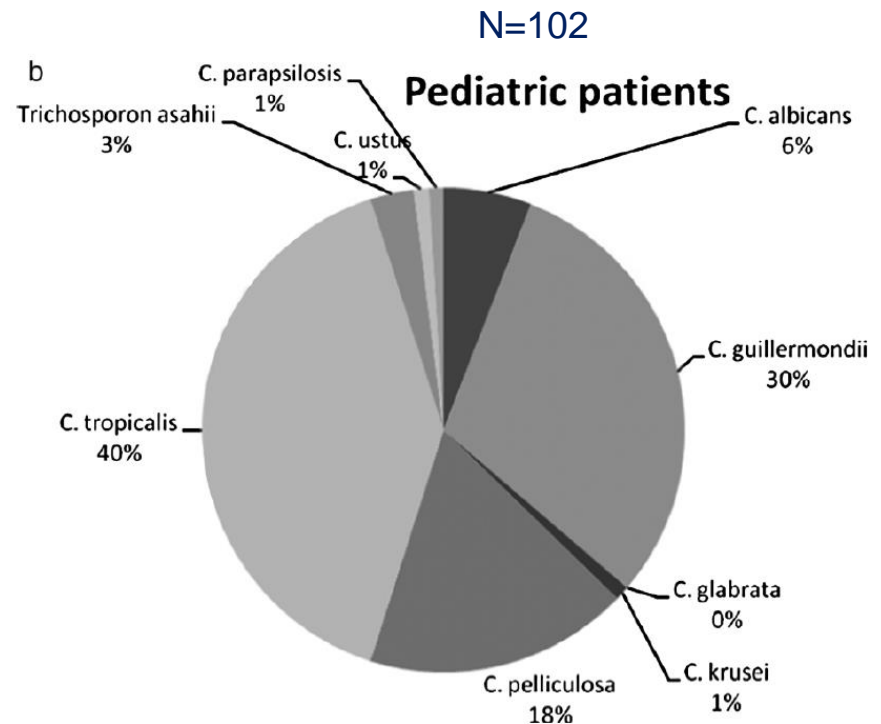
# *C. pelliculosa*

4-month prospective study in a tertiary care center, India, Sept-Dec 2007



- *C. pelliculosa*, also known as *Pichia anomala* or *Hansenula anomala*, is mainly found in plants, fruits and oil.
- Ten out of 14 episodes in this cohort were reported from a single hospital, clustered in the ICUs. Tan et al. Clin Microbiol Infect 2015;21:946
- In the preceding year, this hospital identified a monoclonal outbreak of *C. pelliculosa*.

J Microbiol Immunol Infect 2013;46:456



Chakrabarti et al. Scand J Infect Dis 2009;41:275-84

*Pichia anomala* (*C. pelliculosa*) outbreak in paediatric wards during 1996-1997.



# Emerging opportunistic yeast infections

- *Candida krusei*<sup>1</sup>
- *Candida guilliermondii*<sup>2</sup>
- *Candida rugosa*<sup>3</sup>
- *Candida lusitanae*<sup>4</sup>
- *Candida dubliniensis*<sup>5</sup>
- *Candida pelliculosa*<sup>6</sup>
- *Candida kefyr*<sup>6,7</sup>
- *Candida nivariensis*<sup>8</sup>
- *Candida norvegensis*
- *Cryptococcus humicolus*
- *Cryptococcus uniguttulatus*
- *Geotrichum capitatum*<sup>9</sup>
- *Hansenula*
- *Saccharomyces cerevisiae*

1. Intrinsic resistance to fluconazole, susceptible to voriconazole
2. Potential for decreased susceptibility to polyenes, azoles, flucytosine, and the echinocandins
3. Cross resistance to fluconazole and voriconazole
4. Can develop secondary resistance to amphotericin B
5. Can develop stable fluconazole resistance, especially in patients with HIV/AIDS
6. Outbreak
7. Dairy products
8. Gardens or potted plants
9. Presence of blastoconidia with hyphae differentiates *Trichosporon* from *Geotrichum*, predominantly in Italy.



# *Candida haemulonii* and Closely Related Species, Korea

- A yeast species that often exhibits **antifungal resistance (AmB, azole)**, rarely causes human infection
- Recovered from 8 patients with fungemia and 15 patients with chronic otitis media in 5 hospitals in Korea during 2004–2006.
- Species identification
  - Vitek 2 YST yeast card system: identified as *C. haemulonii*
  - API 20C system: identified as *Kodamaea ohmeri* and *Rhodotorula glutinis*
- Drug resistance were associated with **therapeutic failure**
- All susceptible to caspofungin and micafungin

# One size not fit all

## Activity of antifungal drugs against emerging yeasts

	Azoles		Polyenes	Echinocandins
	Fluconazole	Voriconazole	Amphotericin formulations	Caspofungin
<i>Candida</i> species				
<i>Candida glabrata</i>	Susceptible (dose dependent) to resistant	Susceptible (dose dependent) to resistant	Susceptible to intermediate susceptibility	Susceptible*
<i>Candida tropicalis</i>	Susceptible	Susceptible	Susceptible	Susceptible*
<i>Candida parapsilosis</i>	Susceptible	Susceptible	Susceptible	Susceptible to resistant*
<i>Candida krusei</i>	Resistant	Susceptible (dose dependent) to resistant	Susceptible to intermediate susceptibility	Susceptible
<i>Candida kefyr</i>	Susceptible	Susceptible	Susceptible	Susceptible
<i>Candida lusitanae</i>	Susceptible	Susceptible	Susceptible to resistant	Susceptible*
<i>Candida dubliniensis</i>	Susceptible to resistant	Susceptible	Susceptible	Susceptible
<i>Candida rugosa</i>	Very low activity	Low activity	Susceptible	Susceptible
<i>Candida quilliermondii</i>	Low activity	Susceptible	Susceptible	Susceptible
<i>Trichosporon</i> species				
<i>Trichosporon asahii</i>	Low activity	Susceptible	Resistant	Resistant
<i>Trichosporon beigelii</i> (cutaneum)	Low activity	Low activity	Resistant	Resistant
<i>Rhodotorula</i> species	Very low activity	Variable susceptibility/ very low activity	Susceptible	Resistant
Non-neoformans cryptococcus species				
Overall	Low activity	Susceptible	Susceptible	NA
<i>Cryptococcus laurentii</i>	Very low activity	NA	Susceptible*	Resistant
Other uncommon yeasts				
<i>Geotrichum</i> species	Variable susceptibility	Susceptible	Susceptible	NA
<i>Hansenula anomala</i>	Fluconazole: low activity; itraconazole: very low activity	Susceptible	Susceptible	Susceptible
<i>Malassezia</i> species	Fluconazole: low activity; itraconazole: susceptible	Susceptible	Variable susceptibility	NA
<i>Saccharomyces</i> species	Low activity/variable susceptibility	Susceptible	Susceptible	NA

Resistant was defined as less than 40% of isolates tested reported as active. Susceptible was defined as more than 90% of isolates tested reported as active. Low activity was defined as 60–89% of isolates tested reported as active. Very low activity was defined as 40–59% of isolates tested reported as active. NA=data not available. \*Susceptible but resistance reported after exposure (ie, breakthrough infections).



# Comparative In vitro Activities of Various Antifungal Drugs against *Candida* and *Cryptococcus* Singapore General Hospital, 2004-2006

Fungal isolates	No.	POS		FLU		VOR		AMB		CAS	
		MIC <sub>50</sub>	MIC <sub>90</sub>	MIC <sub>50</sub>	MIC <sub>90</sub>	MIC <sub>50</sub>	MIC <sub>90</sub>	MIC <sub>50</sub>	MIC <sub>90</sub>	MIC <sub>50</sub>	MIC <sub>90</sub>
All <i>Candida</i> sp.	100	0.064	1.5	0.38	16	0.032	0.38	0.38	0.75	0.094	0.25
<i>C. albicans</i>	24	0.032	0.064	0.125	0.19	0.006	0.008	0.094	0.125	0.032	0.125
<i>C. tropicalis</i>	28	0.094	0.125	0.38	0.75	0.047	0.094	0.5	1	0.094	0.19
<i>C. glabrata</i>	27	1	2	16	48	0.25	0.5	0.5	0.75	0.094	0.19
<i>C. parapsilosis</i>	12	0.023	0.047	0.38	0.75	0.012	0.023	0.125	0.5	0.25	1
<i>C. dubliniensis</i>	7	0.012	0.023	0.094	0.25	0.004	0.006	0.012	0.032	0.125	0.125
<i>C. krusei</i>	1	0.25	0.25	24	24	0.19	0.19	0.5	0.5	0.25	0.25
<i>C. famata</i>	1	0.006	0.006	0.094	0.094	0.003	0.003	0.016	0.016	0.19	0.19
<i>Cryptococcus</i> sp.	10	0.125	0.38	8	32	0.023	0.094	0.25	0.25	>32	>32
<i>C. neoformans</i>	8	0.125	0.38	4	32	0.016	0.094	0.25	0.38	>32	>32
<i>C. gattii</i>	2	0.19	0.5	8	32	0.064	0.125	0.25	0.25	>32	>32

Etest, MIC in µg/mL

Tan AL et al. Ann Acad Med Singapore 2008;37:841

# Antifungal Susceptibility of 216 cryptococcal clinical isolates in Taiwan, 1997–2010

Antifungal agent	Genotype	No. of isolates	Minimum inhibitory concentration (μg/mL)					% (No.) above ECV	
			Range	Geometric Mean	MIC <sub>50</sub>	MIC <sub>90</sub>	ECV	This study	Global studies <sup>a</sup>
Amphotericin B									
	VNI	203	0.03–1	0.48	0.5	0.5	0.5	3.4% (7)	2.8%
	VNII	4	0.13–1	0.42	0.5	1	NA <sup>a</sup>		
	VGI	3	0.25–0.25	0.25	0.25	0.25	0.5	0%	0.8%
	VGII	6	0.06–1	0.31	0.5	1	1	0%	0.8%
Flucytosine									
	VNI	203	0.13–32	1.14	1	2	8	0.5% (1)	3.4%
	VNII	4	0.13–2	0.30	0.19	2	NA <sup>a</sup>		
	VGI	3	0.5–1	0.63	0.5	1	4	0%	4.3%
	VGII	6	1–2	1.59	2	2	16	0%	2.9%
Fluconazole									
	VNI	203	0.03–16	2.35	4	8	8	0.5% (1)	2.9%
	VNII	4	0.13–8	0.84	0.75	8	NA <sup>a</sup>		
	VGI	3	1–4	2	2	4	8	0%	1.2%
	VGII	6	0.13–16	5.04	8	16	32	0%	6.9%
Voriconazole									
	VNI	203	0.03–0.25	0.06	0.06	0.13	0.25	0%	2.4%
	VNII	4	0.03–0.13	0.05	0.05	0.13	NA <sup>a</sup>		
	VGI	3	0.03–0.06	0.04	0.03	0.06	0.5	0%	0%
	VGII	6	0.13–0.25	0.20	0.25	0.25	0.25	0%	4.1%



**Is antifungal resistance in  
*Aspergillus* and other molds a  
problem in Asia?**

---

# Comparative In vitro Activities of Various Antifungal Drugs against Moulds

Fungal isolates	No.	POS		ITR		VOR		AMB		CAS	
		MIC <sub>50</sub>	MIC <sub>90</sub>	MIC <sub>50</sub>	MIC <sub>90</sub>	MIC <sub>50</sub>	MIC <sub>90</sub>	MIC <sub>50</sub>	MIC <sub>90</sub>	MIC <sub>50</sub>	MIC <sub>90</sub>
All moulds	50	0.047	>32	0.19	>32	0.094	4	0.25	3	0.016	>32
<i>Aspergillus sp.</i>	34	0.023	0.125	0.094	0.25	0.064	0.19	0.125	2	0.008	0.047
<i>A. fumigatus</i>	12	0.032	0.047	0.094	0.19	0.094	0.125	0.19	0.25	0.003	0.047
<i>A. niger</i>	12	0.012	0.032	0.064	0.19	0.032	0.064	0.125	0.19	0.012	0.064
<i>A. flavus</i>	7	0.094	0.19	0.19	0.25	0.125	0.19	2	4	0.008	0.023
<i>A. clavatus/A. nidulans</i>	3	0.016	1	0.25	0.38	0.094	0.25	0.032	0.38	0.016	0.032
<i>Fusarium solani</i>	10	>32	>32	>32	>32	2	4	3	4	>32	>32
Others	6	0.38	>32	1	>32	6	64	0.25	>32	>32	>32

Etest, MIC in µg/mL

Tan AL et al. Ann Acad Med Singapore 2008;37:841



# Invasive infections caused by moulds other than *Aspergillus*

- A tertiary care hospital in Taiwan, 2000-2008
- 103 patients with cultures positive for non-*Aspergillus* moulds
- The overall mortality rate was 40.7%, and was highest in cases zygomycosis.

Characteristic	<i>Fusarium</i>	<i>Paecilomyces</i>	<i>Zygomycetes</i>	<i>Scedosporium</i>
Number (%) of patients	12 (44.4)	7 (25.9)	5 (18.5)	3 (11.1)
Age, median (range)	39.5 (0–79)	60 (9–82)	37 (23–81)	65.4 (48–78)
Male	6 (50)	4 (57)	3 (60)	3 (75)
Diagnosis				
Acute leukaemia	5	2	4	0
Lymphoma	1	0	0	0
MDS	1	0	0	0
Solid tumour	1	1	0	1
AIDS	0	2	0	0
Diabetes mellitus	0	2	0	0
HSCT	0	0	1	0
Solid organ transplant	1	0	1	1
Immunological risk factors				
Steroid use	6	1	1	2
Neutropenia < 500/ $\mu$ L	4	2	2	0
Lymphopenia < 1500/ $\mu$ L	7	2	2	0

# Cutaneous T cell lymphoma with acute leukemic change



**CR3**



**Relapse 3**



**Disseminated fusariosis**





# Recommendations for treatment of *Fusarium* infection in immunocompromised patients

Population	Intention	SoR	QoE	Comment
Immunocompromised patients	First-line treatment Voriconazole	A	Ilt,r	Therapeutic drug monitoring required Response rate was associated with underlying condition and infection site
	Liposomal amphotericin B	B	Ilt,r	Fungi may be resistant to amphotericin B
	Amphotericin B lipid complex	C	III	Limited case reports
	Amphotericin B deoxycholate	D	Ilt,u	Fungi often resistant to amphotericin B Breakthrough infections may occur Excessive toxicity
	Any echinocandin	D	III	Intrinsically resistant
	Any combination therapy	C	III	Limited reports Combination not better than voriconazole alone
	Salvage treatment Posaconazole	A	II	Overall success rate 50% Breakthrough infections Therapeutic drug monitoring required
	Voriconazole	A	III	Substantial efficacy Therapeutic drug monitoring required

QoE, quality of evidence; SoR, strength of recommendation.

# Get ready for voting...

Go to webpage: [pollev.com/mmtn](http://pollev.com/mmtn)

Voting question on the next slide

Wifi network: MMTN2016  
Password: 2016mmtn



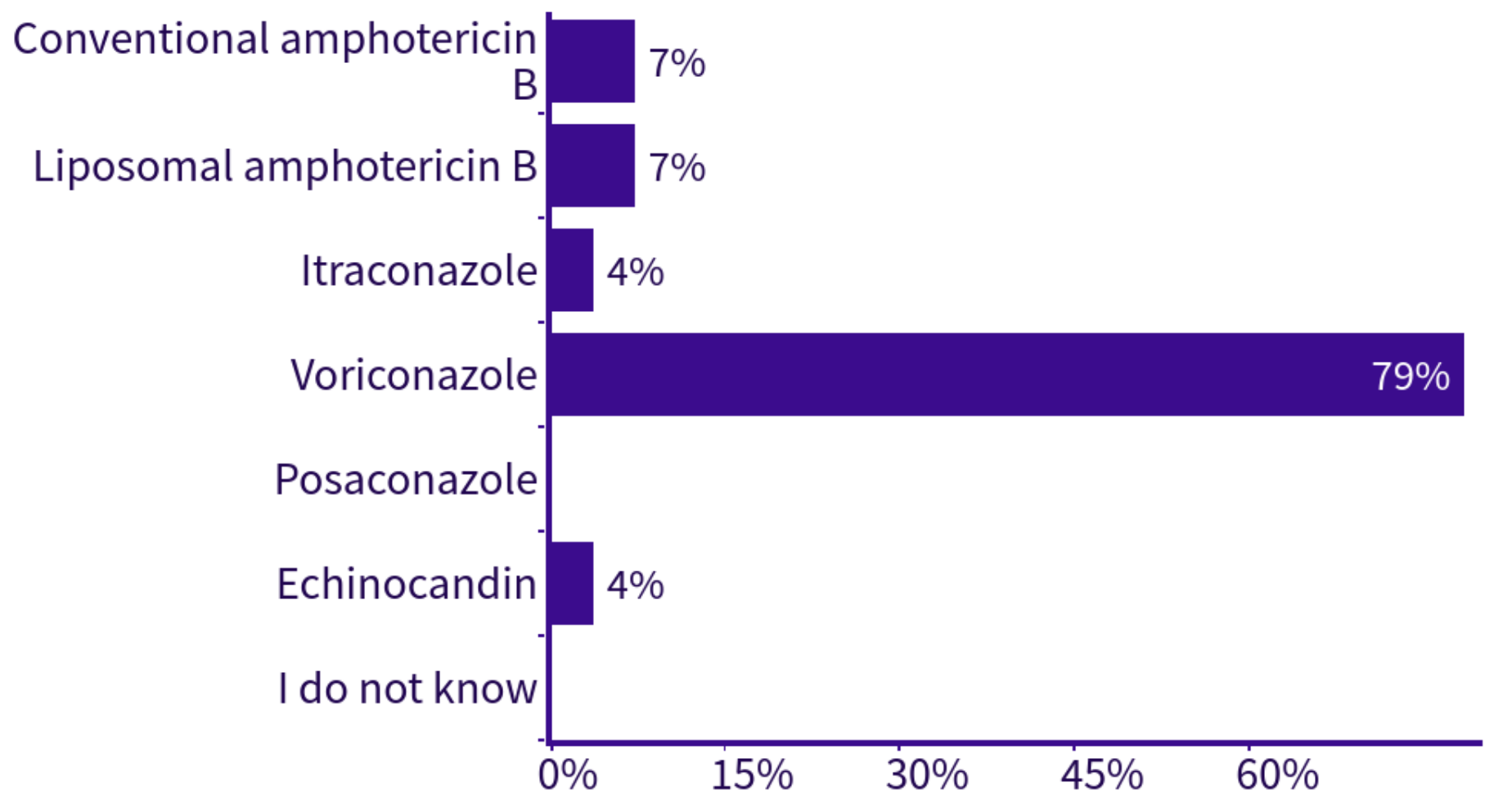
Q. Which is your most commonly used antifungal agent for invasive aspergillosis?

---

- A. Conventional amphotericin B
- B. Liposomal amphotericin B
- C. Itraconazole
- D. Voriconazole
- E. Posaconazole
- F. Echinocandin
- G. I do not know

# Which is your most commonly used antifungal agent for invasive aspergillosis?

 Poll is full and no longer accepting responses



Total Results: 28



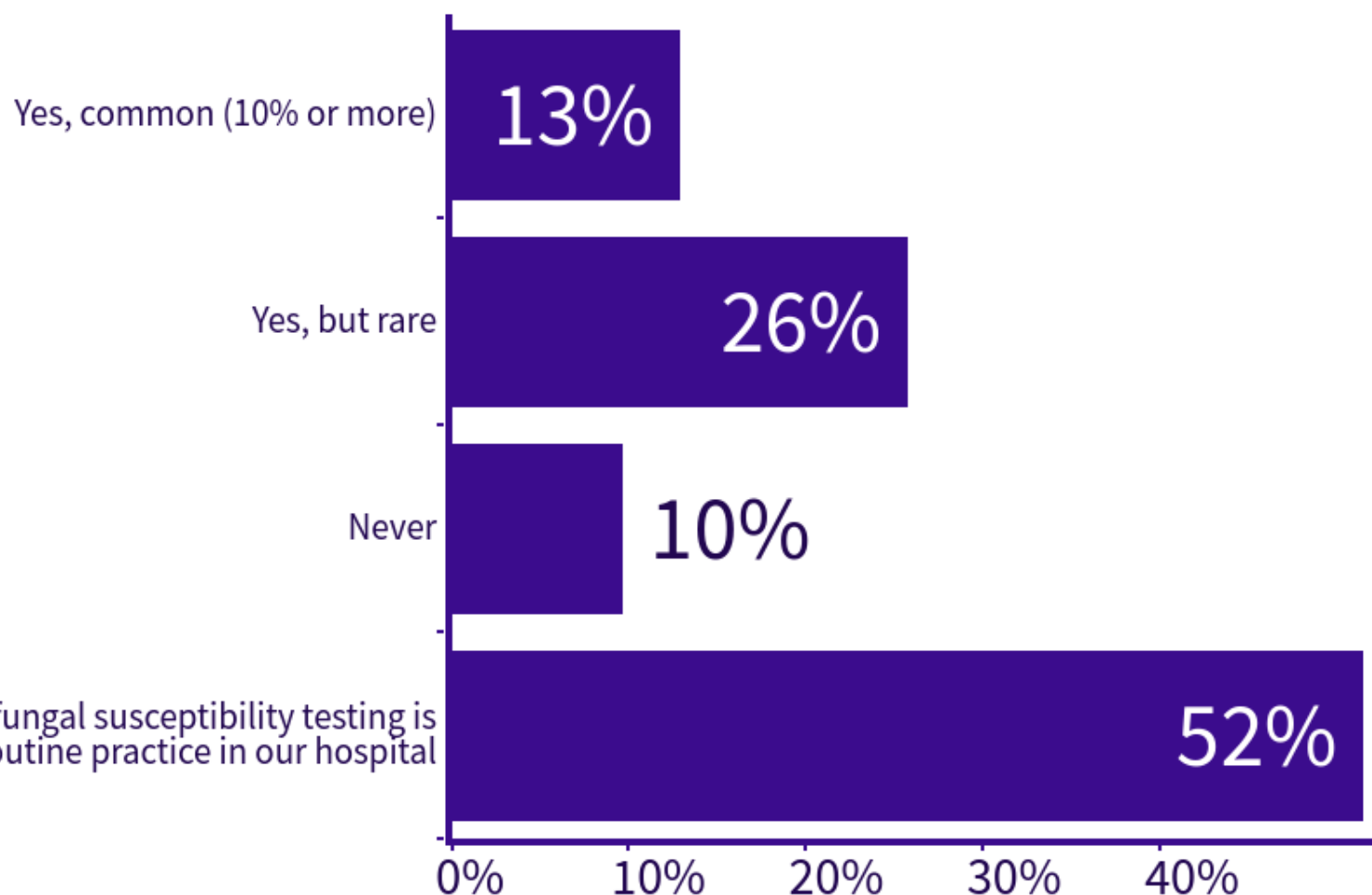
Q. Does antifungal resistance compromise the use of current antifungal agent for invasive aspergillosis in your hospital?

---

- A. Yes, common (10% or more)
- B. Yes, but rare
- C. Never
- D. I do not know as antifungal susceptibility testing is not available for routine practice in our hospital

# Does antifungal resistance compromise the use of current antifungal agent for invasive aspergillosis in your hospital?

**i** Poll is full and no longer accepting responses



Total Results: 31



# Mutations in the *cyp51A* gene and MICs of azole antifungals in 6 *A. fumigatus* isolates isolated from a patient with lung aspergilloma

## Itraconazole

No

Yes 6M

No 2M

Yes

Yes

Yes

Isolate	Mutation in <i>cyp51A</i>	MIC (mg/L)	
		itraconazole	voriconazole
AF1	–	0.25	0.25
AF2	M220I	>16	1
AF3	–	0.5	0.5
AF4	G54R	>16	0.5
AF5	G54R	>16	0.5
AF6	G54R	>16	0.5

# Recovery of TR34/L98H and TR46/Y121F/T289A Resistance Mechanisms in *Aspergillus fumigatus*

TR<sub>34</sub>/L98H

TR<sub>46</sub>/Y121F/T289A

Country	First Case	Type of Isolate	Year of Publication [Reference]	Country	First Case	Type of Isolate	Year of Publication [Reference]
Netherlands	1998	C + E	2008 [13]	United States	2008	C	2015 [25]
Italy	1998	C + E	2015 [23]	Netherlands	2009	C + E	2013 [24]
Turkey	2000	C	2015 [26]	Belgium	2012	C + E	2012 [12, 27]
Spain	2003	C	2013 [12, 28]	Germany	2012	C + E	2015 [12, 15]
Australia	2004	C	2015 [29]	India	2012	E	2014 [30]
Iran	2005	C + E	2013 [12, 30]	France	2013	C + E	2015 [31]
Belgium	2006	C + E	2012 [12, 27]	Tanzania	2013	E	2014 [32]
Denmark	2007	C + E	2010/2011 [12, 33]	Denmark	2014	C	2015 [33]
China	2008–2009	C	2011 [30, 34]	Spain	2014	C	2015 [28]
India	2008	C + E	2012 [12, 30]	Colombia	2015	E	2015 [35]
United Kingdom	2009–2011	C + E	2009 [12, 36]				
France	2010	C + E	2012 [12]				
United States	2010	C	2015 [25]				
Germany	2012	C + E	2012 [12, 15]				
Taiwan	2011	C	2015 [37]				
Kuwait	2013	C + E	2015 [30, 38]				
Poland	2006–2014	C	2015 [39]				
Colombia	2015	E	2015 [35]				

Abbreviations: C, clinical; E, environmental.

<sup>a</sup> Due to space restriction, we were not able to include all individual publications. We have cited reviews, which included reports from individual countries over the years.



# Azole-resistant *Aspergillus fumigatus* isolates carrying TR<sub>34</sub>/L98H mutations, Taiwan

The prevalence rates of azole resistance

- global 3–6%
- Taiwan 6.5%

Patient, sex/age(y)	Isolate	Sample type	Underlying diseases	Minimum inhibitory concentrations, mg/L					
				ITZ	VCZ	POS	AMB	TBZ	PEN
Azole-resistant isolate, n=3			two azole-naïve patients						
1, M/59	A31	BAL	Lung cancer	≥16 (≥16 <sup>b</sup> )	4 (4 <sup>b</sup> )	1 (0.5 <sup>b</sup> )	0.5	32	>32
2, F/66	B44	sputum	chronic hepatitis C, cirrhosis of liver, diabetes without control and adrenal insufficiency	>16 (≥16 <sup>b</sup> )	2 (1 <sup>b</sup> )	1 (0.5 <sup>b</sup> )	0.5	16	>32
	B51	sputum		>16 (≥16 <sup>b</sup> )	2 (1 <sup>b</sup> )	1 (0.5 <sup>b</sup> )	0.5	16	>32
Azole-susceptible isolates, n=35				0.06/0.12 [0.03-0.25] b,c	0.25/0.5 [0.12-1] b,c	0.03/0.06 [0.015-0.06] b,c	-	1/2 [0.5-4] <sup>c</sup>	2/2 [0.5-4] <sup>c</sup>

# **Environmental Multiple-Triazole-Resistant *A. fumigatus* Strains Carrying the TR34/L98H Mutations in the *cyp51A* Gene in India**

---

- A total of 44 (7%) *A. fumigatus* isolates from 24 environmental samples were triazole resistant.
- Cross-resistance to voriconazole, posaconazole, itraconazole and to six triazole fungicides used extensively in agriculture.
- In contrast to the genetic uniformity of azole-resistant strains the azole-susceptible isolates from patients and environments in India were genetically very diverse.
- All Indian environmental and clinical azole resistant isolates shared the same multilocus microsatellite genotype



# **The possible mechanisms or sources of antifungal resistant fungal pathogens**

---

1. De novo occurrence of mutation in causality pathogens following the use of antifungal agents
2. Selection of drug resistant fungal pathogens following the use of antifungal agents
3. Cross transmission of antifungal resistant fungal pathogens from other patients or environment in the healthcare settings
4. Acquisition of antifungal resistant fungal pathogens from the agricultural environment

# Limitation of current published data

---

- **Definition**

- Clinical breakpoints

- **Detection**

- Identification to species levels
- In vitro susceptibility testing in routine laboratories

- **Publication**



# Conclusions

---

- Antifungal resistance has emerged and spread in Asia.
- Antifungal susceptibility vary by fungal pathogens and by region/country/hospital
- Population surveillance data to guide local practice (empirical therapy)
- Detection of fungal pathogen and identification to species level to guide definitive therapy
- Evaluate in vitro susceptibility of individual isolate for selected fungi and/or patients